



# IN VIVO ANTITUMOR ACTIVITY OF HERBIMYCIN A, A TYROSINE KINASE INHIBITOR, TARGETED AGAINST BCR/ABL ONCOPROTEIN IN MICE BEARING BCR/ABL-TRANSFECTED CELLS

Mihiro Okabe,\* Yoshimasa Uehara,† Takayuki Noshima,‡ Toshiyuki Itaya,\*  
Yasuyuki Kunieda,\* and Mitsutoshi Kurosawa\*

\*The Third Department of Internal Medicine; †The Second Department of Pathology, Hokkaido University School of Medicine, Sapporo 060, Japan; and ‡The Department of Bioactive Molecules, National Institute of Health, Tokyo 162, Japan

(Received 7 March 1994. Revision accepted 1 July 1994)

**Abstract**—Herbimycin A, a benzoquinoid ansamycin antibiotic, has been shown to reverse the oncogenic phenotype of p60<sup>v-src</sup> transformed cells because of the inhibition of src protein tyrosine kinase. We previously demonstrated that herbimycin A displayed antitumor activity on the *in vitro* growth of Philadelphia chromosome-positive leukemia cells and BCR/ABL-transfected murine hematopoietic FDC-P2 cells through the inhibition of BCR/ABL protein tyrosine kinase. In this study, the transformed FDC-P2 cells were demonstrated to be tumorigenic in syngeneic DBA/2 mice. The intraperitoneal (i.p.) injection of the transformed tumor cells into DBA/2 mice induced infiltrations of abdominal organs, and then all of the mice died within time periods proportional to the cell numbers of inoculation. In mice that received an i.p. inoculation with greater than  $1 \times 10^5$  cells, *in vivo* administration of herbimycin A by i.p. injection inhibited tumor formation and significantly prolonged survival time, and further, in mice inoculated with  $1 \times 10^4$  cells, herbimycin A completely suppressed the *in vivo* growth of transformant FDC-P2 cells and brought about a complete remission. The present study revealed the *in vivo* efficacy of herbimycin A in mice bearing BCR/ABL-transfected cells.

**Key words:** Herbimycin A, BCR/ABL oncoprotein, protein tyrosine kinase, tyrosine kinase inhibitor, *in vivo* antitumor activity.

## Introduction

The *bcr/abl* oncogene is the product of the Philadelphia chromosome (Ph<sup>1</sup>), resulting from the reciprocal translocation between chromosomes 9 and 22 which are found in Ph<sup>1</sup>-positive leukemias which mainly constitutes more than 90% of chronic myelogenous leukemia (CML) and approximately 20% of adult acute lymphocytic leukemia (ALL) [1-5]. The *bcr/abl* fusion proteins generating from the hybrid *bcr/abl* genes, P210<sup>bcr/abl</sup> and P190<sup>bcr/abl</sup> exhibit the deregulated tyrosine kinase activity of ABL and oncogenic activity *in vitro* and *in vivo* [6-9].

Recent progress in the study of the *bcr/abl* gene and its product in Ph<sup>1</sup>-positive leukemia offers an opportunity for developing several strategies for specific therapy targeted against BCR/ABL [10-14]. Herbimycin A, an inhibitor of protein tyrosine kinase (PTK) that we isolated on the basis of its ability to

cause rat kidney cells transformed by v-src to revert to normal morphology with a loss of PTK activity of the transforming protein of p60<sup>v-src</sup> [15], was shown to display a relatively selective antitumor activity for transformed cells by oncogenes coding PTK [16]. In fact, we demonstrated that herbimycin A displayed a preferential antitumor activity on the *in vitro* growth of Ph<sup>1</sup>-positive cells by reducing BCR/ABL PTK [12]. In order to determine whether or not herbimycin A may offer a new therapeutic potential for Ph<sup>1</sup>-positive leukemia, we have investigated the antitumor activity of *in vivo* administration of herbimycin A in syngeneic DBA/2 mice inoculated with BCR/ABL oncoprotein-transformed murine FDC-P2 cells. Here, we report the *in vivo* efficacy of herbimycin A on mice bearing BCR/ABL oncoprotein-associated tumors.

## Materials and Methods

### Murine hematopoietic cell lines

The murine IL-3 dependent hematopoietic cell line FDC-P2 [17] was used in this study. As control cells, P815.

Correspondence to: Mihiro Okabe, M.D., Third Department of Internal Medicine, Hokkaido University School of Medicine, Kita-15, Nishi-7, Kita-ku, Sapporo 060, Japan (Tel: (011) 716-1161, Ext. 5920; Fax: (011) 706-7867).

mastcytoma cell line, and P388 lymphoma cell line, were used.

#### *Transfection of retroviral vector expressing P210<sup>bcr/abl</sup> into IL-3-dependent FDC-P2 cells*

Five million FDC-P2 cells were cultured by the addition of a mixture of 30  $\mu$ l Lipofectin (Gibco, Berlin, Germany) and 4  $\mu$ g of pGD210 retroviral vectors expressing P210<sup>bcr/abl</sup> oncoprotein [8] for 16 h in serum-free Opti-MEM (Gibco) in the presence of recombinant murine IL-3 (rmIL-3) which was prepared from COS-1 cells transfected with pSV2neo containing mIL-3 cDNA. The cells were selected in a RPMI-1640 medium to which 10% fetal bovine serum (FBS) and 600  $\mu$ g/ml geneticin (G418 sulfate; Gibco) were added in the presence of rmIL-3 for 10 days, and were then further selected in a medium lacking rmIL-3 for IL-3-independent growth. In the absence of mIL-3, parental or mock pZIPNeoSV(X)-transfected FDC-P2 cells died within 72 h, but G418-resistant pGD210<sup>bcr/abl</sup>-transfected cells were able to grow autonomously in a medium which lacked mIL-3. As previously described in detail [12], transformed FDC-P2 cells expressed *bcr/abl* and *neo* mRNAs and the P210<sup>bcr/abl</sup> oncoprotein.

#### *In vitro growth testing for herbimycin A*

The assay of cellular proliferation was carried out by using the colorimetric MTT method. The cells ( $2 \times 10^4$ /well) were seeded in triplicate in 100  $\mu$ l of 10% FBS-added RPMI 1640 with or without herbimycin A, and cultured for 3–5 days in a humidified atmosphere with 5% CO<sub>2</sub> at 37°C. After the addition of 10  $\mu$ l of a 5 mg/ml stock solution of MTT (3-(4,5-dimethylthiazol-2,5-diphenyl tetrazolium bromide) (Sigma Co. Ltd., Munich, FRG) into each well for the last 4 h, formazan crystals were solubilized in 100  $\mu$ l of isopropanol-0.04 N HCl, and then the plates were measured on a MTP-100 microplate reader (Corona Electric, Tokyo, Japan) using a test wavelength of 570 nm with a reference wavelength of 630 nm.

#### *Western blotting and immune complex kinase assay of BCR/ABL*

Transformant FDC-P2 cells were cultured for 16 h in the presence or absence of 0.2  $\mu$ g/ml of herbimycin A, where cell viabilities of both groups were more than 90%. The cells were lysed in the presence of kinase and phosphatase inhibitors, as previously described by Longo [18]. The lysate was clarified by centrifugation at 4°C, and 20  $\mu$ g of proteins in the supernatant were separated by SDS-PAGE in reducing conditions (100 mM dithiothreitol). Proteins were blotted onto a polyvinylidene difluoride (Millipore) membrane. The BCR/ABL oncoprotein was identified using a monoclonal anti-ABL protein (Ab-3, Oncogene Science, Mineola, NY).

The immune complex kinase assay was carried out by using a modified method of that described by Kurzrock et al. [19, 20]. In brief, the cells ( $1 \times 10^7$ ) were harvested, washed with phosphate-buffered saline (PBS), and then lysed at 4°C by sonication in 50 mM Tris-HCl pH 7.5, containing 0.15 mM NaCl, 0.1% SDS, 1% Triton X-100, 0.5% sodium deoxycholate, 1 mM PMSF. The diluted sample was treated with mouse monoclonal antibody against c-*abl*, Ab-3 (Oncogene Science), for 1 h at room temperature and was incubated overnight at 4°C. Immune complexes with anti-ABL antibody were precipitated with mouse anti-IgG antibody-coated protein A (Zymed Laboratories, San Francisco, CA), and immunoprecipitates

were then reacted with 5  $\mu$ Ci of ( $\gamma$ -<sup>32</sup>P) ATP for 10 min on ice. Next, immune complexes were analyzed on 7.5% SDS-polyacrylamide gels, followed by autoradiography with an intensifying screen.

#### *Animals*

Seven-week-old female DBA/2 mice were supplied by Oriental Kobo Co. Ltd (Tokyo, Japan).

#### *Examination of tumor cells and tissues*

Tissues were processed for histological examination and for Southern blot analysis. High molecular DNAs derived from tissues and tumors were digested with restriction enzymes, electrophoresed through 0.8% agarose gel, and then transferred to nylon membranes. The hybridization with a human 3'-*bcr* genomic DNA probe (*bcr*-1; Oncogene Science) was carried out by the method previously described [21].

#### *Administration of herbimycin A*

Herbimycin A was isolated as described previously [15], and was dissolved in phosphate-buffered saline containing 1% (V/V) of Tween-20. Various concentrations of herbimycin A were then administered by i.p. injections daily from days 2–6 and from day 12 to day 16 after either transformed FDC-P2 cells, P518 cells or P388 cells at cell numbers of  $1 \times 10^4$ – $1 \times 10^6$  were inoculated intraperitoneally into syngeneic DBA/2 mice on the first day.

#### *Expression of results*

Unless otherwise indicated, mean values  $\pm$  one S.D. for measurements from triplicate culture have been presented. Bars in figures indicate one S.D. The statistical analysis was carried out using a Student's *t*-test, and results were interpreted to be significantly different when  $p < 0.001$ .

## Results

#### *In vitro antitumor effect of herbimycin A on BCR/ABL-transformed murine cells*

We investigated the *in vitro* effect of herbimycin A against P210<sup>bcr/abl</sup> oncoprotein-associated autonomous growth of FDC-P2 cells transformed by a transfection of the *bcr/abl* gene. Herbimycin A showed no significant inhibition of the growth of parental FDC-P2 cells in the presence of the culture supernatant of WEHI cells containing mIL-3, but did display an inhibitory effect on the growth of transformant FDC-P2 cells at doses showing no toxicity on murine P815 cells and P388 cells (Fig. 1).

#### *Effect of herbimycin A on BCR/ABL oncoprotein and tyrosine kinase*

We previously reported that the antitumor activity of herbimycin A on Ph<sup>+</sup>-positive leukemia cells seems to be associated with an inhibition on the activity of BCR/ABL PTK in spite of having no effect on the expression of BCR/ABL oncoprotein [12]. Similarly, the immune complex kinase assay of BCR/ABL revealed that herbimycin A dramatically suppressed

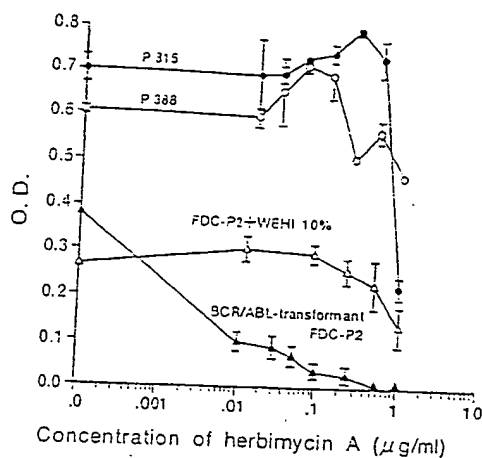


Fig. 1. Effects of herbimycin A on the *in vitro* growth of parental FDC-P2 cells, BCR/ABL-transfected FDC-P2 cells and syngeneic murine hematopoietic malignant cells. Effects of herbimycin A on IL-3-dependent growth of parental FDC-P2 cells in the presence of the culture supernatant obtained from WEHI cells and autonomous growth of transformant FDC-P2 cells by a transfection with BCR/ABL were assayed by MTT proliferation assay after culturing for 72 h. In P815 and P388 cells, MTT proliferation assay was carried out after the 72 h and 5 day cultures, respectively. The growth of P815 cells and P388 cells was not significantly inhibited at low doses of herbimycin A showing an inhibitory effect on the growth of transformant FDC-P2 cells in the 72 h and 5 day cultures. The results of 5 day cultures of P815 and P388 cells are shown.

the autophosphorylation activity of P210<sup>bcf/abl</sup> tyrosine kinase in BCR/ABL-transformed FDC-P2 cells, while it did not affect the amount of BCR/ABL oncoprotein (Fig. 2).

#### Mice bearing tumor cells transfected with BCR/ABL

Transformant FDC-P2 cells were transplantable by either i.p. or i.v. injection into syngeneic DBA/2 mice, whereas parental FDC-P2 cells were not transplanted. Mice inoculated with transformant FDC-P2 cells by i.p. injections exhibited a gradual increase of abdominal tumors and died within 5–11 weeks post-inoculation. Postmortem examinations revealed that tumor cells formed masses on the abdominal walls and infiltrated mainly into mesenteric and retroperitoneal lymph nodes (Fig. 3(A)). The invasions were observed in the liver, spleen, pancreas and peritoneum (Fig. 3(A)), but were not noted in other organs including the lungs and kidneys. The presence of the *bcr/abl* gene in tumors was confirmed by Southern blot analysis with a human *bcr* probe (Fig. 3(B)).

#### In vivo efficacy of herbimycin A in mice bearing BCR/ABL oncoprotein-associated tumors

The study regarding determination of lethal dose

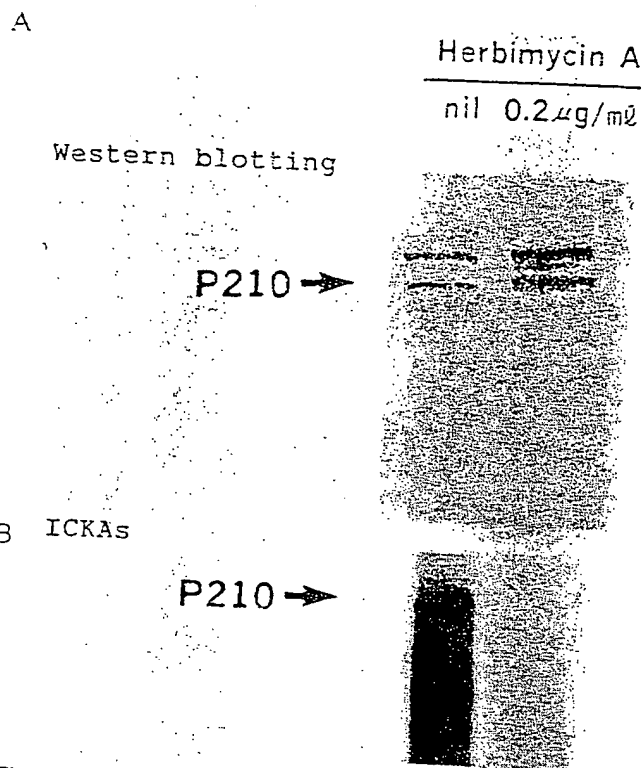


Fig. 2. Suppression of BCR/ABL PTK activity by herbimycin. (A) For the western blotting of BCR/ABL oncoprotein, transformant FDC-P2 cells were cultured for 16 h in the presence or absence of 0.2 µg/ml of herbimycin A, and then cellular protein was isolated. Twenty micrograms of protein were subjected to western blotting. (B) For the immune complex kinase assay (ICKAs), transformant FDC-P2 cells were treated with or without 0.2 µg/ml of herbimycin for 16 h. After this the cell extracts were immunoprecipitated with anti-*c-abl* monoclonal antibody. The resulting immune complexes were analyzed for their autophosphorylation activities.

in DBA/2 mice has revealed that LD<sub>50</sub> of a single i.p. injection of herbimycin A is approximately 20 mg/kg body weight. The intraperitoneal injection of herbimycin daily for 5 days at less than 2.0 mg/kg body weight was well tolerated with no obvious toxicity. A marked reduction in subsequent tumor formation and a prolongation in the survival rates were evident following herbimycin treatment at doses of 1.0–2.0 mg/kg body weight in mice inoculated with greater than  $1 \times 10^5$  of transformant cells (Fig. 4(A)). Furthermore, the i.p. administration with herbimycin A at 2.0 mg/kg body weight completely protected against tumor formation and brought about cures in all mice inoculated with  $1 \times 10^4$  cells, whereas the i.p. administration at 1.0 mg/kg protected against tumor formation in two out of five mice (Fig. 4(B)). In contrast, P815 cells and P388 cells, which were not sensitive to herbimycin A *in vitro*, were transplantable into syngeneic DBA/2 mice, and all mice inoculated i.p. with amounts of greater than  $1 \times 10^4$

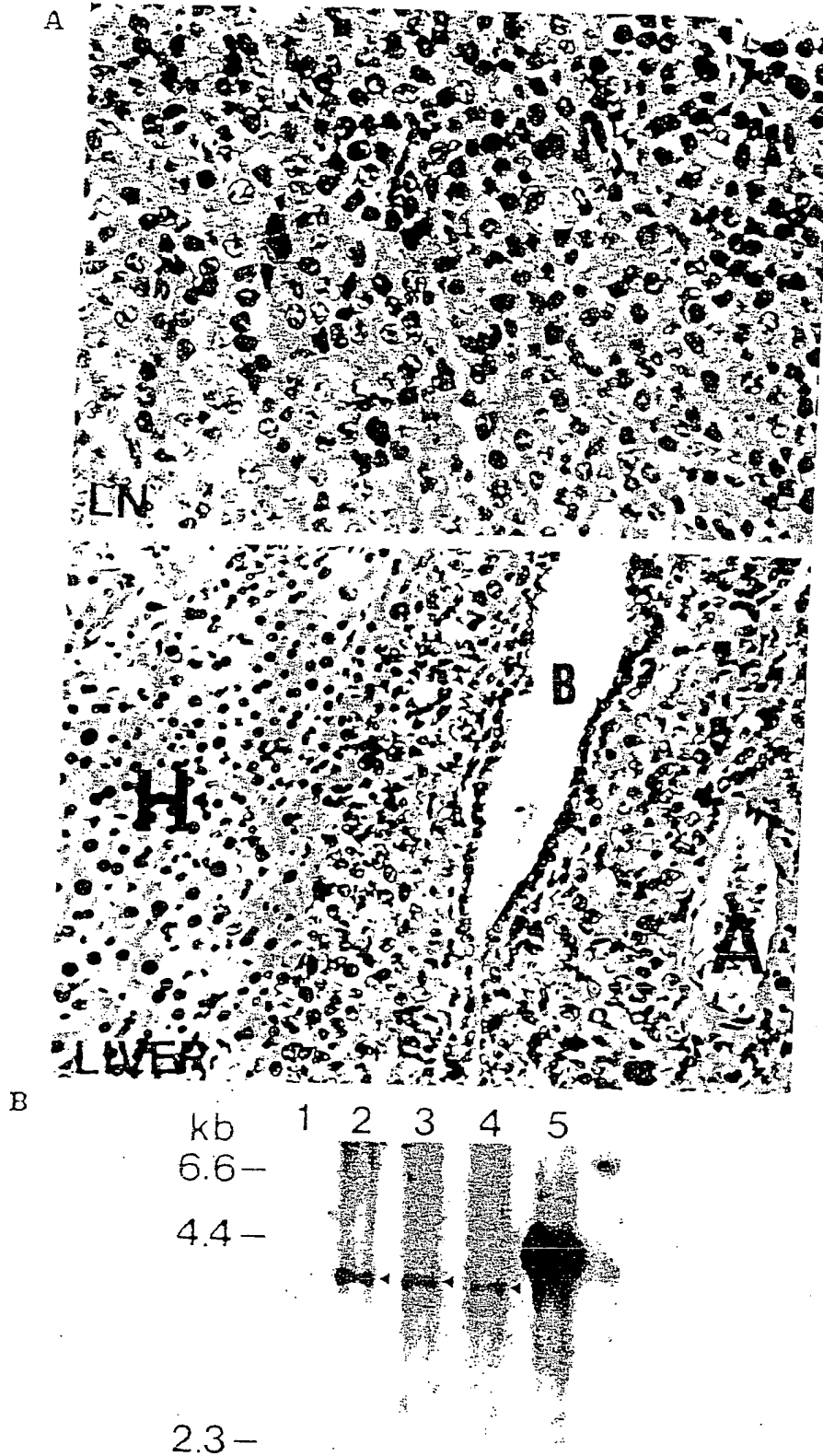


Fig. 3. Histological examination of tumors. (A) Pathological findings of the lymph nodes and the liver. Micrograph of the retroperitoneal lymph node (LN) showing the replacement by tumor cells. Small polygonal cells with prominent nuclei are evident (HE,  $\times 350$ ). Micrograph of the liver: tumor cells are observed throughout the portal tract (HE,  $\times 200$ ). B, bile duct; A, hepatic artery; H, normal hepatocytes. (B) Southern blot analysis revealed the presence of the *bcr/abl* gene. High-molecular weight DNAs isolated from tissues were digested with *HindIII* and were subjected to the Southern blot using 3' M-*bcr* genomic DNA probe. Lane 1, parental FDC-P2 cells; lane 2, transformed FDC-P2 cells; lane 3, lymph nodes; lane 4, liver; lane 5, human placenta DNA. Arrows indicate the signals derived from *bcr/abl* gene.

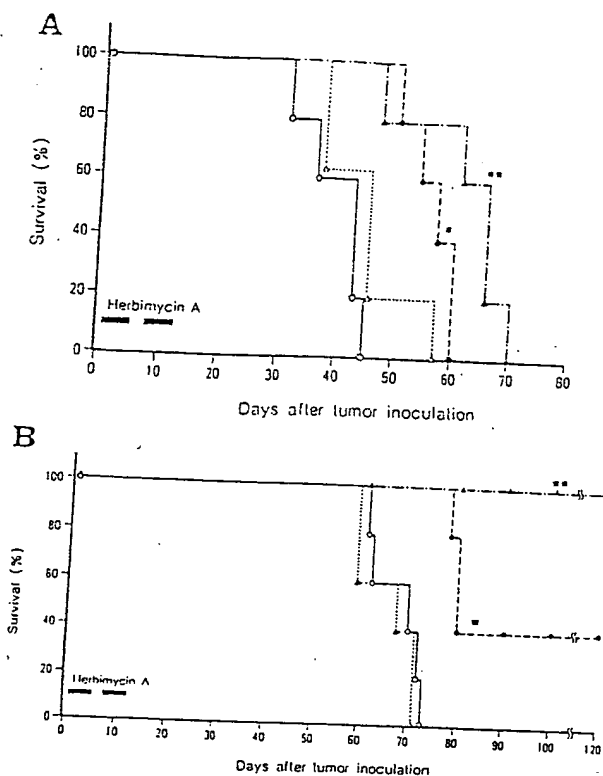


Fig. 4. Therapeutic effects of herbimycin A on DBA/2 mice inoculated with BCR/ABL-transformed FDC-P2 cells. Five DBA/2 mice per group were administered i.p. with herbimycin A (0.1 mg/kg body weight,  $\Delta$ --- $\Delta$ ; 1.0 mg/kg body weight,  $\bullet$ --- $\bullet$ ; 2.0 mg/kg body weight,  $\blacktriangle$ --- $\blacktriangle$ ) on days 1-5 and 8-12 after i.p. inoculation of  $1 \times 10^6$  tumor cells (A) or  $1 \times 10^4$  tumor cells (B). Control groups (O-O) were administered with vehicle only. ■, ■■: significant,  $p < 0.001$  (Student's  $t$ -test).

cells died within 12 weeks. In this case, herbimycin A administration had no effect on their survival rates.

### Discussion

PTK activity has been demonstrated to be associated with stimulatory effects of many growth factors and with transforming oncogenes such as *src*, *abl*, *ros*, *fps*, *yes*, and *fes*. These findings indicate that a variety of PTK may play a critical role in the proliferation and differentiation of cells [22] as well as in some malignant transformations of cells, where  $\text{Ph}^1$ -positive leukemia is representative in human malignancies. Until now, several tyrosine kinase inhibitors including genistein, erbstatin and lavendustin have been developed on the basis of inhibition of EGF-associated tyrosine kinase. In contrast to these inhibitors of receptor-type tyrosine kinase,

herbimycin A, which was isolated from the cultured filtrate of *Streptomyces* sp. MH237-CF-8, was capable of causing a reversion of the transformed Rous sarcoma virus-infected rat kidney cell to normal morphology at modest concentrations via reduction of  $\text{p60}^{\text{src}}$  PTK activity [15]. Furthermore, we previously demonstrated that herbimycin A preferentially suppressed  $\text{Ph}^1$ -positive leukemia cell growth and BCR/ABL oncoprotein-associated transformed murine cells via the inhibition of BCR/ABL PTK activity [12].

In this study, we evaluated the *in vivo* efficacy of herbimycin A in a murine model bearing BCR/ABL oncoprotein-expressing transformed cells. The transformed FDC-P2 cells by a transfection of BCR/ABL grew autonomously *in vitro* and were tumorigenic in syngeneic DBA/2 mice, in which the i.p. inoculation of transformant FDC-P2 cells showed the induction of massive swelling of abdominal lymph nodes and the infiltration of abdominal organs and brought about fatal outcomes in mice. In this murine model bearing the BCR/ABL-associated tumors, the i.p. administration of herbimycin A at concentrations of less than 2 mg/kg body weight for 5 days consecutively for 2 weeks significantly prolonged the survival times of the mice when they were inoculated with more than  $1 \times 10^5$  tumor cells and, furthermore, completely protected against *in vivo* tumor formation in mice that received  $1 \times 10^4$  cells with no overt side effects. Similarly, Honma has previously described the *in vivo* antitumor effect of herbimycin A in the murine model inoculated with highly *v-abl*-expressing murine myeloid cell lines, CI cells [23]. Recently, it has been shown that herbimycin A also is a potent cytotoxin against malignant human tumors with primitive neural features, and the tumorigenicity in nude mice of these sensitive cell lines can be markedly reduced by systemic or topical administration of herbimycin A without any apparent toxicity to the whole animal [24].

More recently, erbstatin and tyrphostins, derivative synthetics of erbstatin, provided an important insight into future therapeutic developments against various types of cancers with amplified ErbB-1 (EGF-R) and ErbB-2 (Neu) through inhibiting PTK activity [25-27]. Furthermore, it has been noted that PTK blockers from the tyrphostin family can discriminate between normal ABL and transforming ABL proteins [28]. In addition to these studies, our development and study of herbimycin A, and further development of chemical modifications might provide an important insight into future therapeutic development according to a new strategy of *bcr/abl* oncoprotein-targeted therapy against  $\text{Ph}^1$ -positive leukemia.

**Acknowledgements**—We are grateful to Dr R. A. van Etten and Dr. G. Q. Daley (Massachusetts Institute of Technology) for providing pGD210 plasmid construct and Dr M. Kobayashi (Hokkaido University School of Medicine) for providing pSV2neo containing murine IL-3 cDNA.

This study is supported in part by a Grant-in-Aid for Scientific Research from the Ministry of Education, Science and Culture of Japan.

## References

- Kurzrock R., Gutterman J. U. & Talpaz M. (1989) The molecular genetics of Philadelphia chromosome-positive leukemias. *New Engl. J. Med.* 319, 990.
- Groffen J., Stephenson J. R., Heistekamp N., Klein A. de, Bartram C. R. & Grosveld G. (1984) Philadelphia chromosomal breakpoints are clustered within a limited region. *bcr*, on chromosome 22. *Cell* 36, 93.
- Hermans A., Heisterkamp N., Lindern M. von, Groffen J., Boostma D. & Grosveld G. (1987) Unique fusion of *bcr* and *abl* genes in Philadelphia chromosome positive acute lymphoblastic leukemia. *Cell* 51, 33.
- Clark L. C., McLaughlin J., Crist W. M., Champlin T. & Witte O. N. (1987) Unique form of the *abl* tyrosine kinase distinguish Ph<sup>1</sup>-positive CML from Ph<sup>1</sup>-positive ALL. *Science* 235, 1985.
- Kurzrock R., Shtarid M., Romero P., Kloetzer W. S., Talpas M., Trujillo J. M., Blick M., Beran M. & Gutterman J. U. (1987) A novel *c-abl* protein product in Philadelphia-positive acute lymphoblastic leukemia. *Nature* 325, 631.
- Daley G. Q. & Baltimore D. (1988) Transformation of an interleukin 3-dependent hematopoietic cell line by the chronic myelogenous leukemia-specific P210<sup>bcr/abl</sup>. *Proc. natn. Acad. Sci. U.S.A.* 85, 9312.
- Lugo T. G., Pendergast A.-M., Muller A. J. & Witte O. N. (1990) Tyrosine kinase activity and transformation potency of *bcr-abl* oncogene products. *Science* 247, 1079.
- Daley G. Q., Van Etten R. A. & Baltimore D. (1990) Induction of chronic myelogenous leukemia in mice by the P210<sup>bcr/abl</sup> gene of the Philadelphia chromosome. *Science* 247, 824.
- Heisterkamp N., Jenster G., Hoeve J. ten, Zovich D., Pattengale P. & Groffen J. (1990) Acute leukemia in BCR/ABL transgenic mice. *Nature* 344, 251.
- Szczylik C., Skorski T., Nicolaides N. C., Manzella L., Malaguarnera L., Venturelli D., Gewirtz A. M., & Calabretta B. (1991) Selective inhibition of leukemia cell proliferation by BCR-ABL antisense oligodeoxynucleotides. *Science* 253, 562.
- Okabe M., Kuni-eda Y., Sugiura T., Tanaka M., Miyagishima T., Saiki I., Minagawa T., Kurosawa M., Itaya T. & Miyazaki T. (1991) Inhibitory effect of interleukin-4 on *in vitro* growth of Ph<sup>1</sup>-positive acute lymphoblastic leukemia. *Blood* 78, 1574.
- Okabe M., Uehara Y., Miyagishima T., Itaya Y., Tanaka M., Kuni-eda Y., Kurosawa M. & Miyazaki T. (1992) Effect of herbimycin A, an antagonist of tyrosine kinase, on BCR/ABL oncoprotein-associated cell proliferation: abrogative effect on the transformation of murine hematopoietic cells by transfection of a retroviral vector expressing oncoprotein P210<sup>210</sup> and preferential inhibition of *in vitro* growth of Ph<sup>1</sup>-positive leukemia cells. *Blood* 80, 1330.
- Dolnick B. J. (1991) Antisense agents in cancer research and therapeutics. *Cancer Invest.* 9, 135.
- Maeriat P., Lewalle P., Taj A. S., Philippe M., Laron-delle Y., Vaerman J. L., Wildmann C., Goldman J. M. & Miahau J. L. (1993) Retrovirally transduced antisense sequences stably suppress P210<sup>bcr/abl</sup> expression and inhibit the proliferation of BCR/ABL-containing cell lines. *Blood* 81, 502.
- Uehara Y., Hori M., Takeuchi T. & Umezawa T. (1985) Screening of agents with converted transformed morphology of Rous sarcoma virus-infected rat kidney cells to normal morphology: identification of an active agent as herbimycin and its inhibition of intracellular *src* kinase. *Jpn. J. Cancer Res.* 76, 672.
- Uehara Y., Murakami Y., Mizuno S. & Kawai S. (1989) Inhibition of transforming activity of tyrosine kinase oncogene by herbimycin A. *Virology* 164, 294.
- Dexter T. M., Garland J., Scott D., Scolnick E. & Metcalf D. (1980) Growth of factor-dependent hemopoietic precursor cell lines. *J. exp. Med.* 152, 1036.
- Longo N., Shuster R. C., Griffin L. D., Langley S. D. & Elsas L. J. (1992) Activation of insulin receptor signaling by a single amino acid substitution in the transmembrane domain. *J. biol. Chem.* 267, 12,416.
- Kurzrock R., Kloetzer W. S., Talpas M., Blick M., Walters R. & Arlinghaus R. B. (1987) Identification of molecular variants of P210<sup>bcr/abl</sup> in chronic myelogenous leukemia. *Blood* 70, 233.
- Okabe M., Kawamura K., Miyagishima T., Itaya T., Goodwyn D., Shoji M., Vogler W. R., Sakurada K., Uehara Y. & Miyazaki T. (1994) Effect of herbimycin A, an inhibitor of tyrosine kinase, on protein tyrosine kinase activity and phosphotyrosyl proteins of Ph<sup>1</sup>-positive leukemia cells. *Leukemia Res.* 18, 213.
- Okabe M., Matsushima S., Morioka M., Kobayashi M., Abe S., Sakurada K., Kakinuma M. & Miyazaki T. (1987) Establishment and characterization of a cell line, TOM-1, derived from a patient with Philadelphia chromosome-positive acute lymphocytic leukemia. *Blood* 69, 990.
- Koch C. A., Anderson D., Moran M. F., Ellis C. & Pawson T. (1993) SH2 and SH3 domains: elements that control interactions of cytoplasmic signaling proteins. *Science* 252, 668.
- Honma Y., Okabe-kado J., Kasukabe T., Hozumi M., Kodama H., Kajigaya S., Suda T. & Miura Y. (1992) Herbimycin A, an inhibitor of tyrosine kinase, prolongs survival of mice inoculated with myeloid leukemia Cl cells with high expression of *v-abl* tyrosine kinase. *Cancer Res.* 52, 4017.
- Whitesell L., Shifrin S. D., Schwab G. & Neckers L. M. (1992) Benzoquinonoid ansamycins possess selective tumoricidal activity unrelated to *src* kinase inhibition. *Cancer Res.* 52, 1721.
- Yaish P., Gazit A., Gilon C. & Levizki A. (1991) Blocking of EGF-dependent cell proliferation by EGF receptor kinase inhibitors. *Science* 242, 933.
- Toi M., Mukaida H., Wada T., Hirabayashi N., Toge T., Hori T. & Umezawa K. (1990) Antineoplastic effect of erbstatin on human mammary and oesophageal tumors in athymic nude mice. *Eur. J. Cancer* 26, 722.
- Yoneda T., Lyall R. M., Alsina M. M., Persons P. E., Spada A. P., Levitzki A., Zilberstein A. & Mundy G. R. (1991) The antiproliferative effects of tyrosine

- kinase inhibitors tyrphostins on a human squamous cell carcinoma *in vitro* and in nude mice. *Cancer Res.* 51, 4430.
28. Anafi M., Gazit A., Gilon C., Ben-Neriah Y. & Lev-

itzki A. (1992) Selective interactions of transforming and normal *abl* proteins with ATP, tyrosine-copolymer substrates, and tyrphostins. *J. biol. Chem.* 267, 4518.